# Visual census methods underestimate density and diversity of cryptic reef fishes 

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The diversity and density of small, benthic reef fishes were estimated using visual census and enclosed rotenone stations. Visual census underestimated the number of species present and the density of common species by up to $91 \%$.
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Key words: sample bias; New Zealand; reef fishes; rotenone; underwater visual census.
Variants of the underwater visual census (UVC) technique first used by Brock (1954) have formed the basis of most studies of reef fish ecology. The non-destructive nature of UVC makes it appealing to workers conducting repeated observations. For study of large reef fishes, the biases associated with UVC have been well documented, whether resulting from variability in fishes (Jennings \& Polunin, 1995; Kulbicki, 1998; Willis et al., 2000) or observer behaviour (Thresher \& Gunn, 1986; Lincoln Smith, 1988; Sale, 1997). However, assemblages of small, cryptic fishes that are strongly associated with the benthos have been either largely ignored, or sampled using UVC with little consideration of methodological bias (Ackerman \& Bellwood, 2000).

As part of a larger research programme, the aim of the present study was to accurately estimate density and assemblage structure of cryptic fishes on subtidal reefs in northeastern New Zealand. To determine whether UVC would be appropriate for such a sampling programme, UVC density estimates were compared with quantitative rotenone samples taken from the exact same area of reef.

Paired sampling of cryptic fishes was done between 15 November 1999 and 10 February 2000 (austral summer) in and adjacent to the Cape Rodney-Okakari Point Marine Reserve in northern New Zealand ( $36^{\circ} 17^{\prime}$ S; $174^{\circ} 48^{\prime}$ E). Each sample consisted of a $3 \times 3 \mathrm{~m}$ plot, which was censused visually by a diver swimming 0.5 m above the substratum (so as not to disturb the fish prior to rotenone sampling) and counting fish in $3 \times 1 \mathrm{~m}$ transects. The same plot was then enclosed by a 1.0 mm mesh, $3 \times 3 \mathrm{~m}$ square cage and sampled using the piscicide rotenone. The cage was composed of a 1 m high wall section, which was weighted at its base with galvanized chain (so it could be moulded to the substratum), and a square roof section. To set the cage up, a 15 kg weight was carefully placed first to define each of the corners of the plot, and two corners of the wall clipped to two adjacent weights. The free corners of the net were then lifted off the bottom by two divers and moved into position. This procedure enabled the walls to be set in place with minimum disturbance to the plot. The roof section was then attached by means of continuous velcro strips. Finally, small floats were attached to the four corners to prevent the net from sagging. Rotenone ( 200 g of $7 \%$ rotenone powder mixed to a paste with sea water) was introduced to the cage either via small gaps at the base (which occurred at high rugosity sites) or through the velcro connection between roof and walls, which was then immediately resealed.

Divers continuously patrolled the circumference up to 2 m from the enclosure for 60 min after the rotenone was released to prevent large fish predators [mostly Parapercis

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Table I. Number of individuals of each species detected in $9 \mathrm{~m}^{2}$ plots ( $n=26$ ) using underwater visual census and quantitative rotenone collections

| Species | Family | Total UVC count | Total rotenone collection |
| :---: | :---: | :---: | :---: |
| Ruanoho whero | Tripterygiidae | 26 | 292 |
| Forsterygion varium | Tripterygiidae | 131 | 236 |
| Forsterygion lapillum | Tripterygiidae | 23 | 90 |
| Dellichthys morelandi | Gobiesocidae |  | 47 |
| Notoclinops segmentatus | Tripterygiidae |  | 22 |
| Gastrocyathus gracilis | Gobiesocidae |  | 21 |
| Pseudophycis breviuscula | Moridae |  | 19 |
| Acanthoclinus marilynae | Plesiopidae |  | 14 |
| Optivus elongatus | Trachichthyidae |  | 13 |
| Trachelochismus melobesia | Gobiesocidae |  | 12 |
| Notoclinus compressus | Tripterygiidae |  | 9 |
| Acanthoclinus rua | Plesiopidae |  | 8 |
| Brosmodorsalis persicinus | Bythitidae |  | 8 |
| Notolabrus celidotus | Labridae | 5 | 8 |
| Scorpaena papillosus | Scorpaenidae |  |  |
| Dermatopsis macrodon | Bythitidae |  | 6 |
| Parika scaber | Monacanthidae |  |  |
| Haplocylix littoreus | Gobiesocidae |  | 4 |
| Conger wilsoni | Congridae |  | 3 |
| Ruanoho decemdigitatus | Tripterygiidae | 1 | 3 |
| Bidenichthys beeblebroxi | Bythitidae |  | 2 |
| Conger verreauxi | Congridae |  | 2 |
| Cryptichthys jojettae | Tripterygiidae |  | 2 |
| Lotella rhacinus | Moridae |  | 2 |
| Parablennius laticlavius | Blenniidae | 5 | 2 |
| Cristiceps aurantiacus | Clinidae |  | 1 |
| Forsterygion malcolmi | Tripterygiidae |  | 1 |
| Gobiopsis atrata | Gobiidae |  | 1 |
| Hippocampus abdominalis | Syngnathidae |  |  |
| Odax pullus | Odacidae |  | 1 |
| Stigmatopora nigra | Syngnathidae |  | 1 |
| Tewara cranwellae | Creediidae |  | 1 |
| Trachelochismus pinnulatus | Gobiesocidae |  | 1 |
| Total |  | 191 | 845 |

colias (Bloch \& Schneider), Pagrus auratus (Bloch \& Schneider), Notolabrus elidotus (Bloch \& Schneider) and Notolabrus fucicola (Richardson)] from accessing gaps at the base, and to capture any fish escaping from the enclosure. One diver then entered the net to complete the collection. The floor of the plot was intensively searched, with all interstices examined so as to ensure a complete census.

A total of 26 plots were sampled within three habitat types (Ecklonia radiata forest, Carpophyllum maschalocarpum stands, and urchin-grazed 'barrens'), which yielded 845 fishes ( 33 species) from rotenone samples, and 191 fishes (six species) from UVC counts (Table I). There was no significant correlation between the two methods in the number of species detected from each plot ( $r^{2}=0 \cdot 014, P>0 \cdot 1 ;$ Fig. 1). Only one species, the crested blenny Parablennius laticlavius (Griffin), was detected more often by UVC than rotenone (Table I). The three most common species [Ruanoho whero Hardy, Forsterygion varium (Bloch \& Schneider) and Forsterygion lapillum Hardy] constituted $94 \%$ and $69 \%$ of UVC and rotenone counts, respectively. The density of all three of these species was underestimated by UVC (Fig. 2). Only $9 \%$ of $R$. whero and $25 \%$ of $F$. lapillum were detected using UVC. Over $55 \%$ of $F$. varium were seen in UVC counts, which agrees with previous characterization of this species as one of the least cryptic in its behaviour (Syms, 1995).


FIG. 1. Relationship between the number of species detected by underwater visual census (UVC) and quantitative rotenone collections. Numbers on the plot indicate a number of superimposed observation points.


FIG. 2. Overall mean density ( $\pm$ s.E.) of the three most common species as estimated by underwater visual census (UVC, $\square$ ) and quantitative rotenone collections ( $\square$ ).

This study supports the conclusions of earlier studies that found UVC to significantly underestimate cryptic fish density (Christensen \& Winterbottom, 1981; Brock, 1982; Kulbicki, 1990; Ackerman \& Bellwood, 2000). The present UVC density estimates might have been biased slightly downward by the need to remain above the substratum (to avoid scaring fish from the plot) while conducting counts. However, even species such as F. varium, thought to be amenable to visual census (Connell \& Jones, 1991), were underestimated by $45 \%$. The use of toxicants or anaesthetics enables detection of species that inhabit reef interstices or burrows and therefore are not usually seen (Kulbicki, 1990; Sayer et al., 1994; Ackerman \& Bellwood, 2000). Although rotenone is destructive, several studies (Willis \& Roberts, 1996; Polivka \& Chotkowski, 1998) have indicated that the effects are generally short-lived, as small reef fishes with high turnover rates recolonize defaunated areas quickly. Accurate estimates of overall reef-fish diversity, abundance, biomass and productivity will require extractive sampling so that the cryptic fishes are not underestimated.

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## References

Ackerman, J. L. \& Bellwood, D. R. (2000). Reef fish assemblages: a re-evaluation using enclosed rotenone stations. Marine Ecology Progress Series 206, 227-237.
Brock, R. E. (1982). A critique of the visual census method for assessing coral reef fish populations. Bulletin of Marine Science 32, 269-276.
Brock, V. E. (1954). A preliminary report on a method of estimating reef fish populations. Journal of Wildlife Management 18, 297-308.
Christensen, M. S. \& Winterbottom, R. (1981). A correction factor for, and its application to, visual censuses of littoral fish. South African Journal of Zoology 16, 73-79.
Connell, S. D. \& Jones, G. P. (1991). The influence of habitat complexity on postrecruitment processes in a temperate reef fish population. Journal of Experimental Marine Biology and Ecology 151, 271-294.
Jennings, S. \& Polunin, N. V. C. (1995). Biased underwater visual census biomass estimates for target-species in tropical reef fisheries. Journal of Fish Biology 47, 733-736.
Kulbicki, M. (1990). Comparisons between rotenone poisonings and visual counts for density and biomass estimates of coral reef fish populations. In Proceedings of the International Society for Reef Studies Congress, Noumea (Ricard, M., ed.), pp. 105-112. Papeete, Tahiti: Université Française du Pacifique.
Kulbicki, M. (1998). How the acquired behaviour of commercial reef fishes may influence the results obtained from visual census. Journal of Experimental Marine Biology and Ecology 222, 11-30.
Lincoln Smith, M. P. (1988). Effects of observer swimming speed on sample counts of temperate rocky reef fish assemblages. Marine Ecology Progress Series 43, 223-231.
Polivka, K. M. \& Chotkowski, M. A. (1998). Recolonization of experimentally defaunated tidepools by northeast Pacific intertidal fishes. Copeia 1998, 456-462.
Sale, P. F. (1997). Visual census of reef fishes: how well do we see what is there? Proceedings of the 8th International Coral Reef Symposium 2, 1435-1440.
Sayer, M. D. J., Cameron, K. S. \& Wilkinson, G. (1994). Fish species found in the rocky sublittoral during winter months as revealed by the underwater application of the anaesthetic quinaldine. Journal of Fish Biology 44, 351-353.
Syms, C. (1995). Multi-scale analysis of habitat association in a guild of blennioid fishes. Marine Ecology Progress Series 125, 31-43.
Thresher, R. E. \& Gunn, J. S. (1986). Comparative analysis of visual census techniques for highly mobile, reef-associated piscivores (Carangidae). Environmental Biology of Fishes 17, 93-116.
Willis, T. J. \& Roberts, C. D. (1996). Recolonisation and recruitment of fishes to intertidal rockpools at Wellington, New Zealand. Environmental Biology of Fishes 47, 329-343.
Willis, T. J., Millar, R. B. \& Babcock, R. C. (2000). Detection of spatial variability in relative density of fishes: comparison of visual census, angling, and baited underwater video. Marine Ecology Progress Series 198, 249-260.

